THYROID AUTOIMMUNITY HAS NO NEGATIVE IMPACT ON INSULIN DYNAMICS IN PREDIABETIC PATIENTS WITH NORMAL THYROID FUNCTION

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Received 13 March 2020, Accepted 03 May 2020

https://doi.org/10.31688/ABMU.2020.55.2.02

ABSTRACT

Introduction. The association of autoimmune thyroid disease (AITD) with diabetes mellitus type-1 (DM1) has been previously documented. However, limited data exist regarding the association of AITD and diabetes mellitus type-2 (DM2).

The objective of the study was to expose the role of AITD on DM2 and the impact of AITD in euthyroid patients with pre-diabetes. Euthyroid prediabetic patients were defined by impaired fasting glucose and/or glucose intolerance. We assessed static and dynamic insulin resistance (IRI) and insulin secretion indices (ISI), and the disposition index (DI).

RéSUMÉ

Introduction. L’association de la maladie thyroïdienne auto-immune (MAIT) avec le diabète de type 1 (DT1) a déjà été documentée. Cependant, des données limitées existent concernant l’association du MAIT et du diabète de type 2 (DT2).

Le but de l’étude était de présenter le rôle de MAIT sur le DT2 et d’explorer l’impact du MAIT chez les patients prédiabétiques ayant une fonction thyroïdienne normale.

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**Results.** Out of 166 patients studied, 31.9% had AITD; impaired fasting glucose (IFG) prevalence was the same with non-AITD patients. In contrast, impaired glucose tolerance (IGT) was less prevalent in AITD compared to non-AITD patients (9.4% versus 28.3%, p=0.008). IRI did not differ between the two groups. The dynamic ISI, 1st phase and 2nd phase insulin release, and the DI showed a worse insulin secretion in non-AITD compared to AITD patients (0.54 versus 0.45, p=0.029; 1029.3 versus 864.3, p=0.033; 354.8 versus 286.5, p=0.035; 0.09 versus 0.07, p=0.02, respectively). On the other hand, hsCRP was higher in AITD versus non-AITD individuals (p=0.008).

**Conclusions.** Although individuals with pre-diabetes and AITD presented with higher levels of low-grade inflammation, their dynamic ISI and the DI, were less impaired.

**Keywords:** insulin resistance indices, QUICKI, HOMA, disposition index, glucose-to-insulin ratio.

**List of abbreviations:**
AITD: autoimmune thyroid disease
Anti-GAD: Antibodies to glutamic acid decarboxylase
BMI: body mass index
DBP: diastolic blood pressure
DI: disposition index
DM: diabetes mellitus
DM1: diabetes mellitus type 1
DM2: diabetes mellitus type 2
GIR: glucose-to-insulin ratio
HbA1c: glycosylated hemoglobin
HOMA: Homeostasis model assessment
HOMA-B: HOMA index of β-cell function
HOMA-IR: insulin resistance HOMA
hs-CRP: high-sensitive C-reactive protein
IGF: impaired fasting glucose
IGT: impaired glucose tolerance
IL: interleukin
incAUCins/glu: incremental area under the insulin to glucose curve
IQR: interquartile range
ISI: insulin secretion index
IRI: insulin resistance index
NHANES III: National Health and Nutrition Examination Survey
OGTT: oral glucose tolerance test
QUICKI: quantitative insulin sensitivity check index
SBP: systolic blood pressure
SD: standard deviation
SI: Static ISI
TNF-α: tumor necrosis factor-α
TPOAb: thyroid peroxidase antibodies
TSH: thyroid stimulating hormone
WHR: waist-to-height ratio
1st PHIS: predicted index of first phase of insulin secretion
2nd PHIS: predicted index of second phase of insulin secretion

**Mots-clés:** indices de résistance à l’insuline, QUICKI, HOMA, indice de disposition, rapport glucose / insuline.
INTRODUCTION

Autoimmune thyroid disease (AITD) and disorders of carbohydrate metabolism are common endocrine disorders in the general population1-2. In a large Nordic unselected population-based study, the prevalence of positive thyroid peroxidase antibodies (TPOAb) was 13.9% in females and 2.8% in males3. Similarly, in another large series of National Health and Nutrition Examination Survey (NHANES III) in the United States population, anti-thyroglobulin antibodies (TgAb) were positive in 10.4±0.5% and TPOAbs, in 11.3±0.4%; positive antibodies increased with age with a clear female preponderance1. On the other hand, a national study from a European country, Spain, revealed that 30% of the population had abnormal carbohydrate metabolism. In particular, the adjusted for age and sex prevalence of established diabetes mellitus type 2 (DM2) was 13.8% (95%CI: 12.8, 14.7%), whereas the prevalence of pre-diabetic states was 3.4% (95%CI: 2.9, 4.0%) for isolated impaired fasting glucose (IFG), 9.2% (95%CI: 8.2, 10.2%) for isolated impaired glucose tolerance (IGT) and 2.2% (95%CI: 1.7, 2.7%) for combined IFG–IGT4. Both DM2 and IGT increased significantly with age, being also more prevalent in men than in women1. Moreover, oral glucose tolerance test (OGTT)-based studies in Europe documented a prevalence of pre-diabetes (including IFG, IGT) ranging from 14% in Spain to 30% in Turkey1.

The association between AITD and alterations in glucose homeostasis has been reported since 19795,6. In 1310 adult persons with DM, 13.4% had thyroid dysfunction (clinical and subclinical hypothyroidism or hyperthyroidism) with the highest prevalence in diabetes type 1 (DM1) (31.4%) and the lowest in DM2 (6.9%)5. The prevalence of thyroid dysfunction among Greek diabetic patients, as defined by the need for thyroxin administration, use of antithyroid drugs, history of thyroidectomy, radioactive iodine treatment, was 12.3%, with women being more frequently affected than men8. In another study, in diabetic patients, the prevalence of thyroid dysfunction defined by the presence of either hypothyroidism or hyperthyroidism (clinical and/or subclinical) was found to be 14.7%, whereas TPOAb were positive in 10.8%9. Inversely, DM2 was present in 27.8% of diabetic patients, the prevalence of thyroid dysfunction or hyperthyroidism (clinical and/or subclinical hypothyroidism or hyperthyroidism) with the highest prevalence among Greek diabetic patients, as defined by serum glucose level at 120 minutes ≥ 140mg/dL (7.77 mmol/L) but < 200 mg/dL (11.1 mmol/L), following a formal 75g OGTT, ii) IFG defined by fasting serum glucose levels ≥100 mg/dL (5.55 mmol/L) but < 126 mg/dL (6.99 mmol/L), according to ADA criteria17. Autoimmune thyroid disease was defined by the presence of TgAb and/or TPOAb antibodies. The population was divided into two groups, patients with AITD and patients without AITD.

All subjects under any medication known to affect glucose metabolism, those with abnormal thyroid function [TSH ≥5 μIU/mL (mIU/L)] defined as subclinical/ clinical hypothyroidism or TSH <0.5 μIU/mL (mIU/L) as subclinical/ clinical hyperthyroidism) and individuals with thyroidectomy were excluded. Pregnant women and individuals with a history of hospitalization during the last 6 months, and hemoglobin levels less than 12 g/dL were also excluded from the study.

The objective of the study was to investigate the possible impact of AITD on insulin secretion and insulin resistance indices (ISI, IRI, respectively), in a population of individuals with prediabetes.

MATERIALS AND METHODS

Subjects

The study included subjects recruited from the Diabetic and Endocrine Outpatient Clinic of the Laiko University Hospital in Athens, Greece. The study was approved by the Scientific Committee of “Laiko” University Hospital (47/14.01.2013). We recruited patients with pre-diabetes, defined by the presence of one of the following criteria: i) IGT defined by serum glucose level at 120 minutes ≥ 140mg/dL (7.77 mmol/L) but < 200 mg/dL (11.1 mmol/L), following a formal 75g OGTT, ii) IFG defined by fasting serum glucose levels ≥100 mg/dL (5.55 mmol/L) but < 126 mg/dL (6.99 mmol/L), according to ADA criteria17. Autoimmune thyroid disease was defined by the presence of TgAb and/or TPOAb antibodies. The population was divided into two groups, patients with AITD and patients without AITD.

Oral glucose tolerance test was performed after 10 hrs overnight fast. Serum glucose (mg/dL) and insulin levels (μIU/mL), were measured at baseline and at 30 min intervals (30’, 60’, 90’, 120’). Glycosylated hemoglobin (HbA1c, %) and high-sensitive C-reactive protein (hs-CRP) were also measured. Plasma glucose, total cholesterol, high density lipoprotein (HDL)-cholesterol, low density lipoprotein (LDL)-cholesterol and triglycerides were measured,
as previously described\textsuperscript{18}. Percent concentration of HbA1c was performed in vitro in whole blood with an immunological method (tholosimetric suppression immuno-analysis, TINIo) in an automatic analyzer of clinical chemistry (Hitachi 912, Roche, France). Serum insulin was measured with the immuno-radiometric assay IRMA (DIA source Immuno-Assays, Louvain-la-Neuve, Belgium). High sensitivity-CRP (mg/L) serum levels were determined by enzyme immunoassay test kit (LI7500, Linear Chemicals, S.L., 08390 Montgat, Barcelona, Spain). The intra- and inter-assay coefficients of variance for hsCRP were 7.5 and 4.1% for low levels and 2.3 and 2.5% for high levels, respectively.

Arterial hypertension was diagnosed according to each individual’s medical and drug history or in the presence of systolic blood pressure (SBP) $\geq 135$mmHg and/or diastolic blood pressure (DBP) $\geq 85$mmHg) \cite{18-21}.

Dyslipidemia was diagnosed based on the LDL levels $\geq 130$mg/dL (3.36 mmol/L) and the administration of hypolipidemic drugs\textsuperscript{22}.

Body weight was measured using analogue scales in light clothing; height was measured barefoot using a stadiometer. Body mass index (BMI, kg/m\textsuperscript{2}) was calculated to assess obesity and waist and waist-to-height ratio (WHR) to assess body fat distribution. Antibodies to glutamic acid decarboxylase (anti-GAD) were randomly measured in the first 25 persons that were enrolled into the study to test for the case of persons with Latent Autoimmune Diabetes of Adults.

### Indices of carbohydrate metabolism – insulin resistance indices (IRI)

**Static IRI**

1/fasting insulin, fasting glucose-to-insulin ratio (GIR), the quantitative insulin sensitivity check index (QUICKI), and insulin resistance Homeostasis model assessment (HOMA) (HOMA-IR) were used to assess insulin action\textsuperscript{23} in the fasting state, using the following formulas:

- **QUICKI** = $1/(\log(\text{fasting insulin (µIU/mL)}) + \log(\text{fasting glucose (mg/dL)}))$\textsuperscript{24}
- **HOMA-IR** = fasting insulin (µIU/mL) $\times$ fasting glucose (mmol/L)/22.5\textsuperscript{25}.

**Dynamic IRI**

The Matsuda index was used to assess dynamic insulin action using the following formula:

Matsuda index$ = (10,000/$\sqrt{\text{fasting glucose} \times \text{fasting insulin}})$ $\times$ ($\text{mean glucose} \times \text{mean insulin during OGTT}$)$^\text{26}$

**β-cell secretion indices (ISI)**

**Static ISI (SISI)**

HOMA index of β-cell function (HOMA-B) was calculated using the following formula:

- **HOMA-B** = $20 \times$ fasting insulin (µIU/mL)/$\text{fasting glucose (mmol/mL)} - 3.5$$^\text{27}$.

**Dynamic ISI**

First phase and second phase insulin secretion and the incremental area under the insulin to glucose curve (incAUCins/glu) were used, by the following formulas\textsuperscript{28}:

- Predicted index of first phase of insulin secretion (1st PHIS) = $1283 + [1.289 \times \text{insulin at 30 minutes (µIU/mL)}] - [138.7 \times \text{glucose at 30 minutes (mmol/mL)}] + [3.772 \times \text{insulin at baseline (µIU/mL)}]$\textsuperscript{29}
- Predicted index of second phase of insulin secretion (2nd PHIS) = $287 + [0.4164 \times \text{insulin at 30 minutes (µIU/mL)}] - [26.07 \times \text{glucose at 30 minutes (mmol/mL)}] + [0.9226 \times \text{insulin at baseline (µIU/mL)}]$\textsuperscript{29}
- **IncAUCins/glu** by the trapezoidal method from 0’ to 120’ min\textsuperscript{31}

**Combined index**

The combined index of insulin action and β-cell secretion is expressed by the disposition index (DI) and the following formula is used:

- **Oral disposition index (DI)** = $\Delta I_{0–30}/\Delta G_{0–30} \times 1/\text{fasting insulin}$\textsuperscript{27}.

The presence of insulin resistance was defined as previously described\textsuperscript{28}: HOMA-IR > 2.16 and/or QUICKI < 0.34 values.

### Assays

The serum TSH levels were measured by a sensitive two-site chemiluminescent immunometric assay with analytical sensitivity: 0.004 µIU/mL, and the coefficient of variation (CV) is less than 5.5% for TSH values comprising between 0.3 and 10µIU/mL. Thyroid antibodies: TgAb < 40 U/mL with analytical sensitivity 20 U/mL with an intra- and inter-assay CV of 3.2% and 4.6%, respectively, and TPOAb < 30 U/mL with analytical sensitivity 10 U/mL with an intra- and inter-assay CV of 5.2% and 3.2%, respectively (IMMULITE 2000 SIEMENS Healthcare Diagnostics Products Ltd. Llanberis, Gwynedd LL55 4EL United Kingdom). High-sensitive CRP (reference
range <5 mg/L) was determined with a highly sensitive latex-based immunoassay. Anti-GAD antibodies were measured by ELISA (Anachem Ltd, Luton, UK).

Statistical analysis
A between-group comparison of categorical variables was carried out by Chi-square test corrected by Fisher’s exact test when appropriate, while for the continuous variables a Student t-test for parametric and Mann-Whitney U test for non-parametric variables was used. Parametric variables are presented as mean value± standard deviation (SD), and non-parametric variables as median values, interquartile range (IQR), minimum-to-maximum values range. A p value<0.05 was considered as statistically significant. SPSS software (SPSS 16. Inc. Chicago, IL) was used for the statistical analysis.

This section may be divided by subheadings. It should provide a concise and precise description of the experimental results, their interpretation as well as the experimental conclusions that can be drawn.

Results

One hundred sixty-six patients with pre-diabetes were recruited; 132 (79.5%) were females. The median age was 50 years and median BMI 32.1 kg/m². Demographic and biochemical characteristics stratified by the presence or absence of AITD are shown in Table 1. The two groups did not differ in terms of age, gender, BMI, waist and waist-to-height ratio (Table 1).

Fifty-three patients (31.9%) had AITD, 34 (64.2%) of those with positive antibodies had positive TgAbs, 50 (94.3%) had positive TPOAbs and 55.8% had both antibodies involved. Seventy (42.2%) of those with positive antibodies had positive antibodies involved. Seventy (42.2%) patients were on thyroxine replacement treatment. All the participants obtained normal thyroid function tests and the serum TSH levels did not differ between the two groups. As expected, more patients with AITD were on thyroxine replacement therapy, compared to non-AITD (p<0.001). One patient (4%) out of 25 had positive anti-GAD antibodies. High sensitivity CRP levels were higher in AITD compared to non-AITD patients, 2.8 (5, 0-16) mg/L versus 1.2 (3, 0-13) mg/L [26.7 (47.6, 0-152.4) nmol/L versus 11.4 (28.6, 0-123.8) mmol/L, p=0.008].

One hundred twenty-nine (92.2%) patients had IFG and 37 (22.3%) had IGT (Table 1). Seventy-seven (46.4%) patients had isolated IFG, 13 (7.8%) had isolated IGT with the remaining 54.2% presenting both abnormalities. IFG prevalence was similar in both groups (AITD: 94.3% versus 91.2%, in non-AITD p=0.55), as opposed to IGT which was less prevalent in AITD (9.4% versus 28.3% in non-AITD, p=0.008).

Fasting serum glucose was similar between AITD and non-AITD individuals [AITD 104 (9, 86-124) mg/dL [5.77 (0.5, 4.77-6.88)] mmol/L; non-AITD 104 (10, 74-125) mg/dL [5.77 (0.56, 4.11-8.55)] mmol/L, p=0.36]. However, 1 hr and 2 hrs post load serum glucose was higher in the non-AITD group, both reaching statistical significance [glucose 60min: AITD 157 (54, 65-227) mg/dL [8.71 (3, 3.61-13.15)] mmol/L; non-AITD 169 (63, 78-275) mg/dL [9.38 (3.5, 4.33-15.26)] mmol/L, p=0.06; glucose 120min: AITD 100 (40, 56-199) mg/dL [5.55 (2.22, 3.11-11.04)] mmol/L; non-AITD 112 (59, 40-199) mg/dL [6.22 (3.27, 2.22-199)] mmol/L, p=0.07]. One hundred twenty-one (72.9%) patients had IR, either by HOMA-IR and/or QUICKI criteria (Table 1).

Static IRI, dynamic IRI and static ISI did not differ between patients with and without AITD. In contrast, dynamic ISI indices, namely IncAUCins/gluc, 1st phase and 2nd phase insulin release, as well as DI, were significantly higher in non-AITD compared to AITD patients (refer to table). DISI indices in AITD individuals were higher irrespective of gender, BMI and HbA1c.

Discussion

The present study provides data suggesting that individuals with pre-diabetes and AITD display a better β-cell insulin secretory profile compared to patients without AITD, for the same level of insulin resistance and despite having a higher level of low-grade chronic inflammation. This finding may simply suggest that thyroid autoimmunity does not confer any additional β-cell secretory dysfunction or, that pre-diabetics with AITD, in order to overcome a similar insulin resistance state as a population without AITD, have to increase their overall β-cell secretion. Since, however, as DI which is a more accurate marker of β-secretory capacity (as it takes into account the level of insulin resistance) was significantly higher in the AITD group, it seems reasonable to assume that β-cell “health” in the AITD group is better than in the non-AITD group. Indeed, this is reflected in the plasma glucose values at 1 hr and 2 hrs post-OGTT, which were lower in individuals with AITD.

Nevertheless, since both clinical and subclinical thyroid disorders have been associated with insulin resistance, in the present study we included patients with normal thyroid function either with or without L-thyroxine replacement treatment to eliminate this confounding factor.

In previous studies, the correlation between DM and thyroid dysfunction has been extensively studied, mostly in regard to DMI, where a causative pathogenic mechanism has been speculated; common
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<table>
<thead>
<tr>
<th>Statical Insulin Resistance Indices (statical IRI)</th>
<th>Pre-diabetes (n=166)</th>
<th>AITD (n=53)</th>
<th>Non-AITD (n=113)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>50 (20, 18-79)</td>
<td>49.5 (20, 18-77)</td>
<td>51 (18, 31-79)</td>
<td>0.43</td>
</tr>
<tr>
<td>Females (%)</td>
<td>1.32 (79.5%)</td>
<td>44 (83%)</td>
<td>88 (77.9%)</td>
<td>0.44</td>
</tr>
<tr>
<td>BMI (kg/m(^2))</td>
<td>32.1 (10, 18-59)</td>
<td>31.4 (12, 18-59)</td>
<td>32.4 (2, 22-58)</td>
<td>0.25</td>
</tr>
<tr>
<td>Waist (cm)</td>
<td>97.5±16.4</td>
<td>98.6±14.8</td>
<td>96.9±17.4</td>
<td>0.58</td>
</tr>
<tr>
<td>Waist-to-height ratio</td>
<td>0.58 (0.13, 0.88)</td>
<td>0.58 (0.15, 0.88)</td>
<td>0.59 (0.11, 0.81)</td>
<td>0.51</td>
</tr>
<tr>
<td>TSH (μIU/mL, mIU/L)</td>
<td>2.3±1.1</td>
<td>2.3±1.0</td>
<td>2.3±1.1</td>
<td>0.065</td>
</tr>
<tr>
<td>Thyroxin replacement (%)</td>
<td>70 (42.2)</td>
<td>31 (58.5)</td>
<td>39 (34.5)</td>
<td>0.004*</td>
</tr>
<tr>
<td>Dyslipidemia</td>
<td>68 (56.2)</td>
<td>22 (51.2)</td>
<td>46 (59)</td>
<td>0.41</td>
</tr>
<tr>
<td>Total Cholesterol (mg/dL, mmol/L)</td>
<td>204.4±38 (5.3±1.0)</td>
<td>208±43.5 (5.4±1.1)</td>
<td>202.4±34.7 (5.2±0.9)</td>
<td>0.48</td>
</tr>
<tr>
<td>HDL (mg/dL)</td>
<td>55.2±15.3 (1.4±0.24)</td>
<td>56.1±16.4 (1.5±0.4)</td>
<td>54.7±14.8 (1.4±0.4)</td>
<td>0.65</td>
</tr>
<tr>
<td>LDL (mg/dL)</td>
<td>127.3±33.1 (3.3±0.9)</td>
<td>131.9±38 (3.4±1.0)</td>
<td>124.7±30 (3.2±0.8)</td>
<td>0.31</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>109.5 (60, 41-409)</td>
<td>110 (51, 41-320)</td>
<td>109 (75, 45-409)</td>
<td>0.04*</td>
</tr>
<tr>
<td>Hypertension</td>
<td>53 (44.2)</td>
<td>3 (7.1)</td>
<td>13 (16.7)</td>
<td>0.17</td>
</tr>
<tr>
<td>SBP (mmHg)</td>
<td>134 (21, 108-166)</td>
<td>135 (21, 108-166)</td>
<td>131 (25, 112-166)</td>
<td>0.18</td>
</tr>
<tr>
<td>DBP (mmHg)</td>
<td>88.2±11.6</td>
<td>86.1±11.2</td>
<td>89±11.8</td>
<td>0.18</td>
</tr>
<tr>
<td>HR (bpm)</td>
<td>76 (21, 43-128)</td>
<td>78 (20, 43-128)</td>
<td>76 (19, 59-112)</td>
<td>0.18</td>
</tr>
<tr>
<td>Family History of Diabetes</td>
<td>54 (46.7)</td>
<td>19 (44.2)</td>
<td>37 (48.1)</td>
<td>0.68</td>
</tr>
<tr>
<td>Ab and/or TPOAb (%)</td>
<td>53 (31.9)</td>
<td>53 (100%)</td>
<td>N/A</td>
<td></td>
</tr>
<tr>
<td>Glucose 0 min (mg/dL, mmol/L)</td>
<td>104 (9, 74-125)</td>
<td>109 (9, 86-124)</td>
<td>104 (10, 74-154)</td>
<td>0.36</td>
</tr>
<tr>
<td>Glucose 60 min (mg/dL, mmol/L)</td>
<td>163 (60, 65-275)</td>
<td>157 (54, 65-237)</td>
<td>169 (63, 78-275)</td>
<td>0.06</td>
</tr>
<tr>
<td>Glucose 120 min (mg/dL, mmol/L)</td>
<td>108.5 (46, 40-199)</td>
<td>100 (40, 56-199)</td>
<td>112 (59, 40-199)</td>
<td>0.07</td>
</tr>
<tr>
<td>Insulin 0 min (μIU/mL, pmol/L)</td>
<td>12.5 (9, 3-34)</td>
<td>12.1 (9, 3-34)</td>
<td>13 (8, 3-28)</td>
<td>0.42</td>
</tr>
<tr>
<td>Insulin 60 min (μIU/mL, pmol/L)</td>
<td>92.2 (82, 15-688)</td>
<td>103.8 (124, 23-598)</td>
<td>81.3 (82, 15-688)</td>
<td>0.59</td>
</tr>
<tr>
<td>Insulin 120 min (μIU/mL, pmol/L)</td>
<td>53.6 (48, 4-405)</td>
<td>55.8 (82, 10-405)</td>
<td>51.8 (64, 4-405)</td>
<td>0.068</td>
</tr>
<tr>
<td>hsCRP (mg/L, nmol/L)</td>
<td>1.7 (0.4-16)</td>
<td>2.8 (0.5-16)</td>
<td>1.2 (0.3-13)</td>
<td>0.008*</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>5.7 (0.5, 4.2-6.5)</td>
<td>5.7 (0.7, 4.2-6.5)</td>
<td>5.7 (0.5, 4.2)</td>
<td>0.58</td>
</tr>
<tr>
<td>IFG (%)</td>
<td>151 (92.2)</td>
<td>50 (94.3)</td>
<td>103 (91.2)</td>
<td>0.55</td>
</tr>
<tr>
<td>IGT (%)</td>
<td>37 (22.3)</td>
<td>5 (9.4)</td>
<td>32 (28.3)</td>
<td>0.004*</td>
</tr>
<tr>
<td>Insulin resistance (HOMA-IR/QUICKI)</td>
<td>121 (72.9%)</td>
<td>41 (77.4%)</td>
<td>80 (70.8%)</td>
<td>0.37</td>
</tr>
<tr>
<td>Glucose/insulin (GIR)</td>
<td>8.5 (3.6-41)</td>
<td>9.2 (7, 3-41)</td>
<td>7.9 (5, 4-32)</td>
<td>0.37</td>
</tr>
<tr>
<td>QUICKI</td>
<td>0.32 (0.04, 0.28-0.42)</td>
<td>0.32 (0.04, 0.28-0.42)</td>
<td>0.32 (0.03, 0.29-0.40)</td>
<td>0.49</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>3.3 (2.5, 0.6-9.5)</td>
<td>3.1 (2.6, 0.6-9.5)</td>
<td>3.5 (2.2, 0.8-7.7)</td>
<td>0.49</td>
</tr>
<tr>
<td>Dynamic Insulin Resistance Index (dynamic IRI)</td>
<td>0.08 (0, 0.03-0.4)</td>
<td>0.08 (0, 0.03-0.4)</td>
<td>0.08 (0, 0.04-0.3)</td>
<td>0.42</td>
</tr>
<tr>
<td>MATSUDA index</td>
<td>3.05 (2.08, 0.72-13)</td>
<td>3.03 (3.03, 0.83-1.3)</td>
<td>2.8 (2.3, 0.72-10.83)</td>
<td>0.31</td>
</tr>
<tr>
<td>HOMA-B</td>
<td>38.9 (27.3, 5.3-124.5)</td>
<td>35.8 (28.6, 5.3-124.5)</td>
<td>42.1 (23.3, 7.8-87.7)</td>
<td>0.37</td>
</tr>
<tr>
<td>Dynamic cell secretion indices (dynamic ISI)</td>
<td>1st phase insulin release</td>
<td>917.6 (909.6, -282.5-2772.5)</td>
<td>864.3 (826.9, -282.5-2772.5)</td>
<td>1029.3 (837.1, 323-2614.2)</td>
</tr>
<tr>
<td>2nd phase insulin release</td>
<td>316.5 (275.6, 24.7-906.3)</td>
<td>286.5 (254.7, 24.7-906.3)</td>
<td>354.8 (268.4, 132.8-851.1)</td>
<td>0.035</td>
</tr>
<tr>
<td>AUC insulin/glucose</td>
<td>0.49 (0.45, 0.09-2.42)</td>
<td>0.45 (0.41, 0.09-2.06)</td>
<td>0.54 (0.49, 0.20-2.42)</td>
<td>0.029</td>
</tr>
<tr>
<td>Combined Insulin Resistance and β-cell secretion index</td>
<td>0.08 (0.07, -0.01-0.55)</td>
<td>0.07 (0.06, -0.01-0.51)</td>
<td>0.09 (0.09, 0.006-0.55)</td>
<td>0.02</td>
</tr>
</tbody>
</table>

*statistical significant value

Footnotes: AUC: area under the curve; BMI: body mass index; DBP: diastolic blood pressure; HDL: high-density lipoprotein; HbA1c: glycoaxylated hemoglobin; HR: heart rate; IFG: impaired fasting glucose; IGT: impaired glucose tolerance; IR: insulin resistance; LDL: low-density lipoprotein; QUICKI: quantitative insulin sensitivity check index; TSH: thyroid stimulating hormone; SBP: systolic blood pressure
susceptibility genes were identified in both autoimmune diseases. An apparent correlation between AITD and DM2 has been speculated in some, but not all the studies, although no pathogenic mechanism has been identified. In a recent study, one in five DM2 subjects had AITD, as opposed to one in twenty seen in control subjects. Although there are studies in agreement with this finding, other studies did not document any association between AITD and DM2, as in the present study. In studies did not document any association between AITD and DM2, as in the present study.

In addition, the presence of metabolic syndrome, with or without obesity, has been shown to be related to AITD in some, but not all studies, while similarly discordant results were published regarding the association of dyslipidemia with AITD. In favor of an association between AITD and cardiovascular risk factors is the fact that AITD has been related to indirect indices of atherosclerosis, such as carotid intima-media thickness.

Several reasons have been put forward to account for the above-mentioned contradictory findings, such as differences in the assays used to determine thyroid auto-antibodies (more sensitive methodologies may result in a higher association), along with inherent variability of the diabetic populations studied, such as duration of diabetes, diabetic control, and medications used. In order to eliminate the impact of the above differences on indices of insulin secretion and/or action (IRI or ISI) and to clarify the relationship of AITD with carbohydrate metabolism abnormalities, we have recruited euthyroid patients with prediabetes not receiving any antidiabetic or insulin-sensitizing medication.

Our data showed that 30% of euthyroid patients with prediabetes have AITD, a percentage that is similar to the 32.9% percentage found in a cohort of non-pregnant patients with DM2 with unknown thyroid function. As a high proportion of patients with prediabetes are likely to develop frank DM, we have speculated that the presence of AITD may confer to prediabetes a higher risk of progression to DM. However, despite the fact that the AITD group exhibited a higher (low-grade) inflammatory state, no difference in insulin resistance between the two groups was observed, while, even further, thyroid autoimmunity was associated with a better β-cell secretory profile, which was reflected in the higher DI, the lower glycemic excursions at 1 hr and 2 hrs post-OGTT and, consequently, in the lower prevalence of IGT in the AITD group. In other words, dysglycemia in individuals with AITD was mainly associated with impaired fasting rather than impaired tolerance of glucose. Clinical studies suggest that the site of insulin resistance varies between IFG and IGT, the first being related mainly with severe hepatic insulin resistance with normal or near-normal muscle insulin resistance, while the latter with only mild hepatic insulin resistance, but with severe insulin resistance at the periphery. AITD may have some impact and inflammatory factors, such as the levels of serum interleukin (IL)-6, tumor necrosis factor-α (TNF-α), IL-12, IL-10, and HOMA-IR, were higher in patients with AITD and hypothyroidism compared to ones with AITD and normal levels of thyroid hormones. On the other hand, we have shown similar TSH levels between groups, suggesting that TSH did not influence our results. Thyroid replacement treatment per se may also influence our findings, since more patients on AITD group were on treatment; however, the subgroups of AITD, with and without replacement treatment, did not differ in any of the parameters studied. Another factor that may be involved is the change in metabolomic patterns and fatty acid metabolism, that may promote insulin resistance. However, the isolated TSH increase, as it was often observed in obesity or euthyroid patients with AITD, is not correlated with insulin metabolism. Slight changes in thyroid hormones may centrally interact with AMP-activated protein kinase (AMPK), decreasing peripheral glucose production and linking glucose regulation to fatty acids synthesis via the carboxylation of acetyl-CoA to form malonyl-CoA, which is catalyzed by acetyl-CoA carboxylase.

Therefore, it may be speculated that the increased low-grade inflammation of the AITD group is preferentially inducing insulin resistance at the hepatic level, rather than the muscle or the adipose tissue level. It cannot be excluded, of course, that our findings may be the result of plain chance. The small sample size, the lack of more specific inflammatory markers, as well as the fact that insulin resistance was not assessed by more sophisticated techniques, do not allow for more robust conclusions to be drawn.

Our study has some limitations that need to be considered. Patients with AITD were studied cross-sectionally, being at various periods of disease evolution and AITD effect on thyroid function. We also recruited patients from the endocrine and diabetic outpatient clinics, who present a source of bias. This explains the higher number of females participating in the study, which could be a potential bias for the assessment of autoimmunity of the population participating in the study. However, despite the small numbers of patients studied in subpopulations analysis, the fact that similar results were seen in normal-weight and lean patients implies that this relationship is real and independent from obesity. Moreover, we found out one patient confirming the previously documented 4% presence of anti-GAD that inclusion or exclusion did not affect the results.
CONCLUSIONS

In conclusion, prediabetic patients with AITD exhibit a better β-cell dependent secretory profile, as assessed by mathematical models associated in spite of an increased inflammatory state. Future larger studies using a prospective design may shed light on the exact mechanism that explains the major impact that thyroid autoimmunity has on insulin-resistance instead of β-cell secretion.

Author Contributions:

K.I.A., G.B. equally contributed to the design of the study, to patients recruitment, analysis of the findings and writing of the manuscript; I.A., L.I.B., P.M., A.K., V.M. contributed to the collection of the data; T.G.P. contributed to data analysis and revision of the manuscript; K.M., L.D. contributed to the design of the study and revision of the manuscript; G.A.K., S.L. equally contributed to the design of the study, to patients recruitment, and revision of the manuscript. All authors read and approved the final manuscript.

Compliance with Ethics Requirements:

“The authors declare no conflict of interest regarding this article”

“The authors declare that all the procedures and experiments of this study respect the ethical standards in the Helsinki Declaration of 1975, as revised in 2008(5), as well as the national law”

“No funding for this study”

Acknowledgments:

None.

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